

European Journal of Cancer 40 (2004) 2604-2612

European Journal of Cancer

www.ejconline.com

Biomarkers of ovarian tumours

Amy V. Rapkiewicz*, Virginia Espina, Emanuel F. Petricoin III, Lance A. Liotta

Laboratory of Pathology, National Cancer Institute/National Institutes of Health, Building 10, Room 2N206, 10 Center Drive, MSC 1500, Bethesda, MD 20892-1500, USA

Received 15 January 2004; accepted 20 May 2004 Available online 10 August 2004

Abstract

Ovarian cancer is one of the most aggressive gynaecological malignancies and most often the high mortality is a direct result of delays in diagnosis. The development of an ovarian cancer-specific biomarker for the early detection of disease has the capacity to improve the dismal survival rate. Currently, there are multiple investigations that are utilising both genomic and proteomic technologies to identify genes, gene products and proteins that may potentially identify diagnostic ovarian cancer biomarkers. Here, we review the studies that are involved in biomarker development for the detection of ovarian cancer.

© 2004 Elsevier Ltd. All rights reserved.

Keywords: Biomarker; Cancer; Microarray; Ovarian; Proteomics

1. Introduction

It is estimated that 25 400 women will be diagnosed with ovarian cancer in 2003 and 14300 will succumb to the disease [1]. The mortality associated with this malignancy begets a huge burden such that it accounts for the greatest number of fatalities among all gynaecological cancers [2]. Unfortunately, the majority of epithelial ovarian cancers remain clinically undetected until patients have developed late stage disease and only a mere 25% of cancers are detected as stage I disease [2]. Once stage III and IV ovarian cancer, which is defined by peritoneal and extra peritoneal metastatic spread, is diagnosed, the survival decreases from 95% at stage I to approximately 20-25% five-year survival despite appropriate treatment [3–6]. Therefore, clinical outcome and possibly survival may be significantly improved by the identification of stage I disease without the need to change surgical or chemotherapeutic approaches. One of the obstacles that is encountered when attempting to identify disease that is confined to the ovary is that affected patients rarely become symptomatic in the early

E-mail address: rapkiewa@mail.nih.gov (A.V. Rapkiewicz).

stages. In contrast to the breast, prostate and colon, the ovary is anatomically more difficult to assess during a routine physical examination and bimanual pelvic examination has not proved to be an effective method of identifying incident cases of early stage disease [7]. Furthermore, non-invasive radiographic assessments alone, such as ultrasonography, frequently cannot distinguish between a benign condition and a malignant lesion, which often results in subsequent costly and unnecessary surgical interventions [8-14]. These difficulties along with the relative rarity of ovarian cancer such that the prevalence in the general population is only 40 cases per 100 000 women older than 50 years, complicates the ability of clinicians to screen for and detect preclinical disease in both premalignant and early stage epithelial ovarian carcinoma.

1.1. Biomarker development

With regards to the early detection and prognostication of ovarian cancer, the need for and development of reliable serum biomarkers, which are both sensitive and specific, remains a long awaited priority. Investigators are aware of this need and the Early Detection Research Network (EDRN) established by the National Cancer Institute have proposed 'guidelines' for the development

^{*}Corresponding author. Tel.: +1-301-594-9577; fax: +1-301-480-9488.

of screening biomarkers [15]. The EDRN delineates a schema with five phases of screening biomarker development, which hopefully will lead to the coordination of various institutions and culminate in the application of a systematic, efficient and effective biomarker-based screening programme. These phases include preclinical exploration, clinical assay and validation, retrospective and prospective screening, as well as conducting a clinical randomised trial for the purpose of assessing the endpoints of cancer screening [15]. When evaluating a biomarker for possible presymptommatic screening, one must consider the low prevalence of ovarian cancer in the general population. Only 1 in 2500 postmenopausal women are affected annually [2]. This number seems rather low in comparison to the 212 600 women that will be diagnosed with breast cancer in that same period of time [National Cancer Institute (NCI). Cancer Net PDQ cancer information summaries, monographs on "screening for breast cancer" (available at http:// www.cancer.gov/cancerinformation/pdq/)]. The specificity of a putative biomarker is challenged by this low incidence. Ovarian cancer is most often definitively diagnosed with a laparotomy, and because this process is costly on many levels, a biomarker with a low specificity cannot be tolerated. A requirement for a specific biomarker has the following statistical profile: a positive predictive value of 100%, a specificity of 99.6% and a sensitivity of 100% [5]. Based on these characteristics, a positive biomarker result will subject ten women to undergo diagnostic exploratory laparotomy or laparoscopy with only one case of ovarian cancer being established.

2. Beyond CA-125

For a widespread screening programme to be successful in reducing the mortality associated with ovarian cancer, recognition of early-stage disease must be targeted. Unfortunately, CA-125, a high molecular weight glycoprotein and serum biomarker [16], does not fit the aforementioned statistical profile, but is approved for monitoring recurrence of disease [4,5,17]. Clinically significant elevations in CA-125 occur in only 80–85% of women diagnosed with early stage disease and 50–60% of late stage disease. Furthermore, CA-125 can also be increased in a variety of other conditions, both benign and malignant, such as the first trimester of pregnancy, breast cancer and endometriosis, as well as lesions that promote any type of peritoneal irritation [18–20]. Recently, several studies have investigated the potential use of CA-125 in combination with other serum markers and/or transvaginal ultrasonography [10,11,21]. Most of these combinations consisted of markers that are highmolecular weight glycoproteins similar to CA-125, such as CA 15-3, CA 72-4, CA 19-9, Lipid-associated sialic

acid (LASA) and OVX1 [22-26]. Some of these biomarker combination strategies have resulted in increased sensitivities at the cost of specificity and, as discussed earlier, this profile is not predictive of disease in screening for early stage ovarian cancer [23,26]. However, one such study focused on evaluating the combination of CA-125, OVX1, LASA and CA72-4 in the setting of a pelvic mass and found that by using a logistic regression analysis, a specificity of 93% could be achieved compared with 76.8% with CA-125 alone [25]. With regards to ultrasonography and CA-125, a study involving more than 22 000 postmenopausal women either over age 45 years or more than 1 year from their last menstrual period, were followed-up for elevated screening levels of CA-125 with transvaginal ultrasonography. Despite significant increases in the median survival between the screened groups from the controls, there was not a significant difference in mortality [5]. There is also evidence that screening for ovarian cancer with serial levels of CA-125 measured over time and interpreted with the risk of ovarian cancer (ROC) calculation may be an optional method to detect preclinical disease [27].

Even though many investigational serum biomarkers such as prostasin and osteopontin [28,29] are emerging as promising answers to this query, new and exciting approaches to cancer diagnosis and screening are on the horizon, such as transcriptional profiling and proteomics [30,31]. Proteomics involves the analysis and illumination of all proteins encoded by the genome. It is these proteins that can be pursued as biomarkers for screening and cancer detection and possibly elucidate the molecular mechanisms involved in oncogenesis.

3. Molecular investigation

3.1. The proteomic horizon

At the NCI–Food and Drug Administration (FDA) Clinical Proteomics Program, a proteomic approach is being investigated for use in the early detection of epithelial ovarian cancer, as well as a variety of other processes such as auto-immune diseases, infectious diseases, prenatal diagnosis, transplantation rejection and vascular disease [32,33]. The proteomic approach has also been investigated for the diagnosis of breast [34], prostate [35] and liver cancers [36]. The complexity and enormity of analysing the proteome is recapitulated in the size and heterogeneity of the proteome itself. Proteins may exist in a variety of modified forms with a conservative estimate of the number of different protein species being greater than 300 000 [37]. Therefore, the identification of appropriate molecular markers, that are both sensitive and specific for epithelial ovarian cancer, is of paramount importance for the NCI-FDA Clinical Proteomics Program. A clinical trial has been proposed which hopefully will lead to the development of the first reliable screening test for ovarian cancer for the general population. Petricoin and colleagues conducted proteinprofiling experiments and generated proteomic spectra by mass spectrometry (surface-enhanced laser desorption and ionisation (SELDI)) [38] and an analytical tool known as a genetic algorithm or artificial intelligence computer algorithm, which discriminates protein patterns into 'diseased', and 'unaffected' training sets (Fig. 1) [38,39]. These experiments are based on the premise that pathological changes in tissue such as the ovary will be reflected in a particular and potentially diagnostic protein 'biosignature' pattern in serum [38-40]. Using this method, low molecular weight serum protein profiling was performed and a specific protein signature was identified that was associated with ovarian cancer from asymptomatic women with a high risk of developing ovarian cancer, such as patients with BRCA1 or BRCA2 mutations, which were provided from the National Ovarian Cancer Early Detection Program (NOCEDP) clinic at Northwestern University Hospital (Chicago, IL, USA) [38,39]. The study comprised analysing and distinguishing a serum protein signature specific for ovarian cancer based on a 'training set' of serum samples from 50 healthy and 50 women with ovarian

cancer (Fig. 2). The bioinformatic program was 'trained' with known mass spectrometry data from women who were unaffected with ovarian cancer and from a cohort of women who were affected with ovarian cancer at various stages of the disease process, from whom blood was obtained prior to surgical or chemotherapeutic intervention. An additional 116 unknown serum samples were used for validation and resulted in a highly accurate classification of the patients without ovarian cancer (95%), as well as those with benign disease (100%), including all stage I cancers. Overall, this result vielded 100% sensitivity and 95% specificity for the identification of ovarian cancer. The positive predictive value for the sample set was 94%, compared with 35% for the CA-125 serum marker for the same samples. This relatively low positive predictive value can be tolerated when screening a high-risk population. However, as discussed, screening the general population requires a positive predictive value that approaches 100%. Notably, all cases of early-stage ovarian cancer were correctly identified based on their signature proteomic pattern. This observation highlights the potential significance of proteomic technology as a diagnostic tool that will play a role in the discovery of new biomarkers for the early detection of disease [41]. The proteomic pattern that was generated for the ovarian cancer serum samples repre-

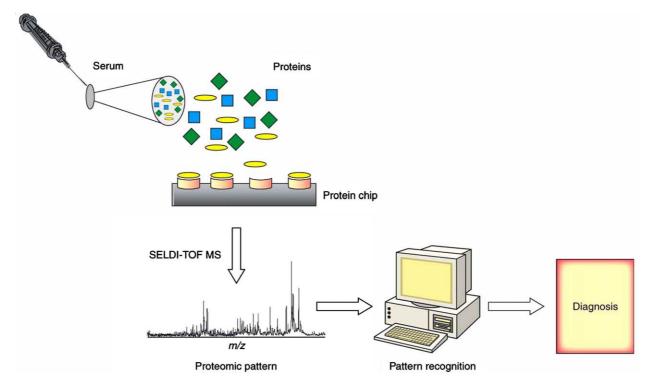


Fig. 1. Disease diagnostics using proteomic patterns. The sample drawn from the patient is applied to a protein chip, which is made up of a specific chromatographic surface. After several washing steps and the application of an energy-absorbing molecule, the species that are retained on the surface of the chip are analysed via mass spectrometry. The pattern of peaks within the spectrum is analysed using sophisticated bioinformatic software to diagnose the source of the biological sample. m/z, mass to charge ratio; SELDI-MS: Surface-enhanced laser desorption/ionisation time-of-flight mass spectrometry. [106] Used with permission.

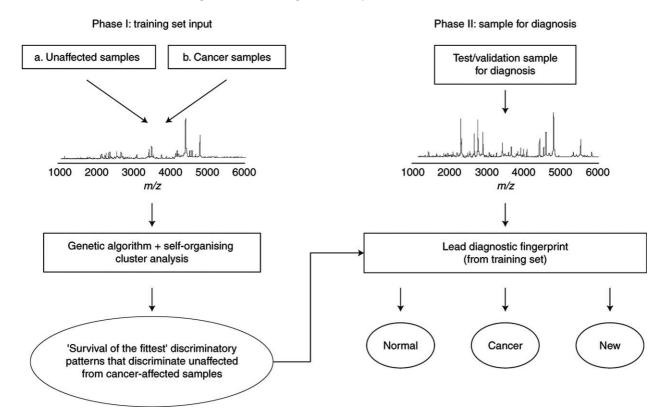


Fig. 2. Bioinformatic analysis of proteomic spectra for the determination of the discriminatory patterns in the training and diagnostic (testing and blind validation) phase [106]. m/z, mass to charge ratio. Used with permission.

sents thousands of proteins and peptides that, to date, remain largely unclassified. However, in other proteomic experiments conducted by the NCI-FDA Tissue Proteomic Initiative on microdissected tissue samples of invasive epithelial ovarian cancer and low malignant potential ovarian tumours, differentially and uniquely expressed proteins were characterised between these two ovarian tumour subtypes [42]. The techniques used to accomplish this experiment were a combination of laser capture microdissection (LCM) and two-dimensional gel electrophoresis (2-D PAGE) [42,43]. Three overexpressed proteins in the invasive cancer samples were selected for further analysis and were identified as glyoxalase I, RhoGDI and FK506 binding protein (FK506BP) [42,43]. All of these proteins have been implicated in oncogenesis affected through various pathways, such as tumour cell apoptosis, DNA synthesis and mutagenesis [44,45]. Specifically, FK506BP and Rho-GDI are indirect mediators of invasion, which is consistent with the pathological differences between the low malignant potential ovarian tumours and invasive ovarian cancer. Furthermore, using the SELDI method of mass spectrometry. Ye and colleagues reported the identification of the haptoglobin- α subunit in the sera of patients with ovarian cancer for use as a possible serum biomarker for the diagnosis of the disease [46]. They reported a significantly higher peak intensity (P = 0.002) in the sera of patients with ovarian cancer compared

with controls, which was validated using an enzymelinked immunosorbent assay (ELISA). Haptoglobin- α subunit may show promise as a serum biomarker used in combination with CA-125 after more studies elucidate the function, as well as the development of a specific antibody assay.

The NCI–FDA studies and others [46,47] demonstrate the feasibility and cost-effectiveness of adding mass spectra analysis not only for diagnosis and the eventual application of this method into routine clinical practice, but also for clinical management of disease such that only a small serum sample is required for analysis [37]. Further clinical trials including large-scale prospective and blinded studies are necessary to validate these initial findings. A future goal is the confirmation of the sensitivity and specificity of this technique as a supplemental tool or an individual approach with regards to the detection of early stage ovarian cancer in the general population.

3.2. The genomic horizon

In general, proteomics represents one of the approaches involved in high-throughput molecular profiling of malignancy and is complemented by genomics. Specifically, DNA microarrays are being used to elucidate the molecular phenotype and gene expression profiles of a number of different cancers, including not

only ovarian, but brain [48,49], breast [50-56], colon [57,58], gastric [59], kidney [60] lung [61–63], prostate [64–67], melanoma [68], small round blue cell tumours of childhood [69] and haematolymphoid system [70–75]. The identification of overexpressed genes or mRNA expression patterns in cancers such as ovarian tumours by cDNA microarray profiling will allow the secreted gene products to be further evaluated as prospective serum biomarkers [76]. Numerous differentially expressed genes in ovarian carcinomas have been reported recently [66,77–84]. However, in terms of their potential usefulness as markers for the early detection of ovarian cancer, the gene product should be secreted. HE4, prostasin, osteopontin, the kallikreins and mesothelin are a sampling of secreted biomarkers that were recognised to be of clinical interest through the use of genomic approaches such as transcriptional profiling.

3.3. The potential genomic markers

The amplification of the HE4 (WFDC2) gene has been shown to occur in ovarian carcinomas [66,85]. The initial identification of the WFDC2 (HE4) was in the epithelial cells of the human epididymis [86], but, subsequently, HE4 overexpression was found in ovarian carcinomas [80,87]. A recent report produced monoclonal antibodies to HE4 epitopes and quantitated the HE4 levels with an ELISA to determine the efficacy of the assay using the sera of postmenopausal women with ovarian cancer compared with matched controls [88]. These blinded studies showed the difference in sensitivity and specificity of the HE4-based ELISA was not statistically significant compared with the CA-125 assay. However, the specificity of HE4 alone was significantly greater (P = 0.001) than CA-125 in it is ability to distinguish between malignant and benign disease. This finding provides incentive for future investigations of HE4 as a diagnostic ovarian cancer biomarker to be used in combination with CA-125.

The gene for prostasin, a serine protease, and osteopontin, a bone morphogen, were identified as upregulated by a cDNA microarray assay using RNA from ovarian cancer cell lines and human ovarian surface epithelial (HOSE) cell cultures [28]. This led to an investigation of the secreted gene products in sera of patients with ovarian cancer. Prostasin, which is normally present in prostatic secretions [89], was shown to be significantly differentially expressed (P < 0.001) in malignant ovarian epithelial cell lines compared with normal HOSE cell lines by real-time quantitative polymerase chain reaction (RT-PCR) [28]. Validation of prostasin expression with anti-prostasin polyclonal antibodies was performed on surgically removed ovarian tissue, both malignant and benign, and did demonstrate more cytoplasmic immunohistochemical staining in the ovarian cancer cells than the normal ovarian tissue. ELISA was then performed on sera from patients in various stages of ovarian cancer and their levels of prostasin were significantly (P < 0.001) higher than in control subjects. Notably, ten patients with stage II ovarian cancer had the highest mean serum levels of prostasin compared with the other stages. This observation is encouraging for the use of prostasin in the early-stage detection of disease. When prostasin and CA-125 serum levels were analysed in combination, it revealed a sensitivity of 92% and a specificity of 94%.

In additional studies performed by the same group, the plasma levels of osteopontin determined by ELISA were reported to be significantly higher (P < 0.001) in the preoperative samples from women with ovarian cancer compared with unaffected individuals, who were either without disease, or had benign pelvic disease or other gynaecological malignancies [29]. While larger validation studies are necessary to further evaluate these markers, the relatively low specificity of prostasin and osteopontin when considering them for the detection of early-stage disease at 94% and 80.4%, respectively, suggests that these biomarkers may prove more useful in a screening panel for the detection of preclinical ovarian cancer.

Similar to prostasin, the human kallikreins are secreted serine proteases [90], and one of the most notable kallikreins, hK3 (prostate-specific antigen) [91,92], is already established as a biomarker for prostate cancer. The gene for human kallikrein 10 (hK10) or normal epithelial cell-specific 1 (NES1), known as KLK10 has been shown to be upregulated in epithelial ovarian cancer [91]. It has been demonstrated that the serum levels of hK10 is significantly elevated (P < 0.001) in presurgical samples from patients with ovarian cancer compared with unaffected individuals [93]. The serum levels of hK10 were not significantly elevated in patients with benign gynaecological disease, which confers a greater specificity compared with CA-125 alone. Presurgical and postsurgical levels of hK10 were also evaluated and considerable reductions were found in approximately 70% of the postsurgical specimens [93]. The human kallikrein gene family contains other potential biomarkers, such as hK6 and hK8, which may have a role in the diagnosis, management and prognostication of ovarian cancer when paired with CA-125 [94,95].

Through the use of oligonucleotide and tissue microarrays, mesothelin gene and protein expression were shown to be significantly elevated in serous carcinomas of the ovary compared with other carcinomas and normal tissue [96]. Validation of this expression was performed using immunohistochemistry and more than 50% of cells in 90% of the serous papillary carcinomas of the ovary were reported as positive. In one study, a soluble member of the mesothelin/megakaryocyte potentiating factor (MPF) family was detected in the sera

of patients with advanced stages of ovarian cancer [97]. Further studies are needed in patients with early stage disease to evaluate the use of soluble mesothelin/MPF in screening for ovarian cancer.

4. Other potential markers

Multiple biomarkers have been considered as candidates for the diagnosis of epithelial ovarian carcinomas [2,98]. However, it becomes evident that these lengthy lists may represent promising future directions, but reproducibility and validation are required for definitive qualification. One such example is the marker is lysophosphatidic acid (LPA). There are conflicting reports with regards to the usefulness of detecting significant differences in the serum levels of LPA in patients with ovarian cancer compared with matched controls when using different modalities for detection [99–101].

5. Discussion

In March 2003, The Helene Harris Memorial Trust Forum on Ovarian Cancer discussed the complex and daunting issues regarding the investigation of biomarkers in ovarian cancer such as universal standardisation of guidelines for comparison and validation studies, challenging data analysis as well as sample availability, both before and after diagnosis [102]. The development of screening biomarkers is further hindered by the lack of understanding regarding the clinical biology and progression of epithelial ovarian cancer from preclinical to metastatic disease [2,78]. Hopefully, the information gathered from clinical trials now underway such as the National Cancer Institute's Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening trial [103] and Bart's 3 trial in the United Kingdom [104], will provide some of this much needed information. The development of ovarian cancer biomarkers is just one example of how the burgeoning field of 'translational medicine' not only has the potential to integrate both clinical proteomics and genomics into daily practice, also but to revolutionise medicine [31,105]. Despite all of these exciting advances, further work is needed to be able to effectively screen and diagnose ovarian cancer.

Conflict of interest

None.

References

- 1. American Cancer Society. Cancer Facts and Figures, 2003.
- Bast Jr RC. Status of tumour markers in ovarian cancer screening. J Clin Oncol 2003, 21(Suppl. 10), 200–205.

- 3. Piver MS, Baker TR, Driscoll DL. Lack of substantial five year disease-free survival by primary aggressive surgery and cisplatin-based chemotherapy or by salvage intraperitoneal cisplatin-based chemotherapy. *Eur J Gynaecol Oncol* 1990, **11**(4), 243–250.
- Ozols R, Rubin S, Thomas G, et al. Epithelial ovarian cancer. Philadelphia, PA, Lippincott Williams and Wilkins, 2000.
- Jacobs IJ, Skates SJ, MacDonald N, Menon U, Rosenthal AN, Davies AP, et al. Screening for ovarian cancer: a pilot randomised controlled trial. *Lancet* 1999, 353(9160), 1207–1210.
- Hoskins P, Eisenhauer E, Vergote I, Dubuc-Lissoir J, Fisher B, Grimshaw R, et al. Phase II feasibility study of sequential couplets of Cisplatin/Topotecan followed by paclitaxel/cisplatin as primary treatment for advanced epithelial ovarian cancer: a National Cancer Institute of Canada Clinical Trials Group Study. J Clin Oncol 2000, 18(24), 4038–4044.
- Grover S, Quinn MA, Weideman P, Koh H, Robinson HP, Rome R, et al. Screening for ovarian cancer using serum CA125 and vaginal examination: report on 2550 females. Int J Gynecol Cancer 1995, 5(4), 291–295.
- Cohen LS, Escobar PF, Scharm C, Glimco B, Fishman DA. Three-dimensional power Doppler ultrasound improves the diagnostic accuracy for ovarian cancer prediction. *Gynecol Oncol* 2001, 82(1), 40–48.
- Campbell S, Bhan V, Royston P, Whitehead MI, Collins WP. Transabdominal ultrasound screening for early ovarian cancer. BMJ 1989, 299(6712), 1363–1367.
- Bromley B, Goodman H, Benacerraf BR. Comparison between sonographic morphology and Doppler waveform for the diagnosis of ovarian malignancy. *Obstet Gynecol* 1994, 83(3), 434-437.
- Woolas RP, Oram DH, Jeyarajah AR, Bast RC, Jacobs IJ. Ovarian cancer identified through screening with serum markers but not by pelvic imaging. *Int J Gynecol Cancer* 1999, 9(6), 497– 501.
- Bourne TH, Campbell S, Reynolds KM, Whitehead MI, Hampson J, Royston P, et al. Screening for early familial ovarian cancer with transvaginal ultrasonography and colour blood flow imaging. BMJ 1993, 306(6884), 1025–1029.
- Menon U, Talaat A, Rosenthal AN, Macdonald ND, Jeyerajah AR, Skates SJ, et al. Performance of ultrasound as a second line test to serum CA125 in ovarian cancer screening. BJOG 2000, 107(2), 165–169.
- Menon U, Talaat A, Jeyarajah AR, Rosenthal AN, MacDonald ND, Skates SJ, et al. Ultrasound assessment of ovarian cancer risk in postmenopausal women with CA125 elevation. Br J Cancer 1999, 80(10), 1644–1647.
- Abstracts of the first scientific workshop of the Early Detection Research Network. September 2000. Dis Markers 2001, 17(1), 3–38
- Bast Jr RC, Klug TL, St John E, Jenison E, Niloff JM, Lazarus H, et al. A radioimmunoassay using a monoclonal antibody to monitor the course of epithelial ovarian cancer. N Engl J Med 1983, 309(15), 883–887.
- Menon U, Jacobs I. Ovarian cancer screening in the general population. *Ultrasound Obstet Gynecol* 2000, 15(5), 350–353.
- Kerbrat P, Lhomme C, Fervers B, Guastalla JP, Thomas L, Tournemaine N, et al. Ovarian cancer. Br J Cancer 2001, 84(Suppl. 2), 18–23.
- Norum LF, Erikstein B, Nustad K. Elevated CA125 in breast cancer—A sign of advanced disease. *Tumour Biol* 2001, 22(4), 223– 228.
- Sjovall K, Nilsson B, Einhorn N. The significance of serum CA 125 elevation in malignant and nonmalignant diseases. *Gynecol Oncol* 2002, 85(1), 175–178.
- Kupesic S, Vujisic S, Kurjak A, Mihaljevic D, Radosevic S. Preoperative assessment of ovarian tumours by CA 125 measurement and transvaginal color Doppler ultrasound. *Acta Med Croatica* 2002, 56(1), 3–10.

- 22. Zhang Z, Barnhill SD, Zhang H, Xu F, Yu Y, Jacobs I, *et al.* Combination of multiple serum markers using an artificial neural network to improve specificity in discriminating malignant from benign pelvic masses. *Gynecol Oncol* 1999, **73**(1), 56–61.
- Woolas RP, Xu FJ, Jacobs IJ, Yu YH, Daly L, Berchuck A, et al. Elevation of multiple serum markers in patients with stage I ovarian cancer. J Natl Cancer Inst 1993, 85(21), 1748–1751.
- 24. Schutter EM, Davelaar EM, van Kamp GJ, Verstraeten RA, Kenemans P, Verheijen RH. The differential diagnostic potential of a panel of tumour markers (CA 125, CA 15-3, and CA 72-4 antigens) in patients with a pelvic mass. *Am J Obstet Gynecol* 2002, 187(2), 385–392.
- Woolas RP, Conaway MR, Xu F, Jacobs IJ, Yu Y, Daly L, et al. Combinations of multiple serum markers are superior to individual assays for discriminating malignant from benign pelvic masses. Gynecol Oncol 1995, 59(1), 111–116.
- van Haaften-Day C, Shen Y, Xu F, Yu Y, Berchuck A, Havrilesky LJ, et al. OVX1, macrophage-colony stimulating factor, and CA-125-II as tumour markers for epithelial ovarian carcinoma: a critical appraisal. Cancer 2001, 92(11), 2837–2844.
- Skates SJ, Menon U, MacDonald N, Rosenthal AN, Oram DH, Knapp RC, et al. Calculation of the risk of ovarian cancer from serial CA-125 values for preclinical detection in postmenopausal women. J Clin Oncol 2003, 21(Suppl. 10), 206–210.
- Mok SC, Chao J, Skates S, Wong K, Yiu GK, Muto MG, et al. Prostasin, a potential serum marker for ovarian cancer: identification through microarray technology. J Natl Cancer Inst 2001, 93(19), 1458–1464.
- Kim JH, Skates SJ, Uede T, Wong Kk KK, Schorge JO, Feltmate CM, et al. Osteopontin as a potential diagnostic biomarker for ovarian cancer. JAMA 2002, 287(13), 1671–1679.
- Bandera CA, Ye B, Mok SC. New technologies for the identification of markers for early detection of ovarian cancer. *Curr Opin Obstet Gynecol* 2003, 15(1), 51–55.
- Mills GB, Bast Jr RC, Srivastava S. Future for ovarian cancer screening: novel markers from emerging technologies of transcriptional profiling and proteomics. *J Natl Cancer Inst* 2001, 93(19), 1437–1439.
- Liotta LA, Kohn EC, Petricoin EF. Clinical proteomics: personalized molecular medicine. *JAMA* 2001, 286(18), 2211–2214.
- Service RF. Proteomics. A sharper focus. Science 2003, 302(5649), 1318
- Li J, Zhang Z, Rosenzweig J, Wang YY, Chan DW. Proteomics and bioinformatics approaches for identification of serum biomarkers to detect breast cancer. Clin Chem 2002, 48(8), 1296– 1304.
- Adam BL, Vlahou A, Semmes OJ, Wright Jr GL. Proteomic approaches to biomarker discovery in prostate and bladder cancers. *Proteomics* 2001, 1(10), 1264–1270.
- 36. Poon TC, Yip TT, Chan AT, Yip C, Yip V, Mok TS, et al. Comprehensive proteomic profiling identifies serum proteomic signatures for detection of hepatocellular carcinoma and its subtypes. Clin Chem 2003, 49(5), 752–760.
- Rosenblatt KP, Bryant-Greenwood P, Killian K, Mehta A, Geho D, Espina V. Serum proteomics in cancer diagnosis and mangement. *Ann Rev Med* 2004, 55, 97–112.
- Petricoin EF, Liotta LA. Mass spectrometry-based diagnostics: the upcoming revolution in disease detection. *Clin Chem* 2003, 49(4), 533–534.
- Petricoin EF, Ardekani AM, Hitt BA, Levine PJ, Fusaro VA, Steinberg SM, et al. Use of proteomic patterns in serum to identify ovarian cancer. Lancet 2002, 359(9306), 572–577.
- Friedrich MJ. Genomics and proteomics may help clinicians individualize cancer treatment. JAMA 2002, 287(22), 2931–2932.
- Liotta LA, Espina V, Mehta AI, Calvert V, Rosenblatt K, Geho D, et al. Protein microarrays: meeting analytical challenges for clinical applications. Cancer Cell 2003, 3(4), 317–325.

- Jones MB, Krutzsch H, Shu H, Zhao Y, Liotta LA, Kohn EC, et al. Proteomic analysis and identification of new biomarkers and therapeutic targets for invasive ovarian cancer. Proteomics 2002, 2(1), 76–84.
- Ardekani AM, Liotta LA, Petricoin III EF. Clinical potential of proteomics in the diagnosis of ovarian cancer. *Expert Rev Mol Diagn* 2002, 2(4), 312–320.
- 44. Olofsson B. Rho Guanine Dissociation Inhibitors: pivotal molecules in cellular signalling. *Cell Signal* 1999, **11**(8), 545–554.
- 45. Takahashi T, Irie RF, Nishinaka Y, Hoon DS. 707-AP peptide recognized by human antibody induces human leukocyte antigen A2-restricted cytotoxic T lymphocyte killing of melanoma. *Clin Cancer Res* 1997, 3(8), 1363–1370.
- 46. Ye B, Cramer DW, Skates SJ, Gygi SP, Pratomo V, Fu L, et al. Haptoglobin-alpha subunit as potential serum biomarker in ovarian cancer: identification and characterization using proteomic profiling and mass spectrometry. Clin Cancer Res 2003, 9(8), 2904–2911.
- 47. Kozak KR, Amneus MW, Pusey SM, Su F, Luong MN, Luong SA, et al. Identification of biomarkers for ovarian cancer using strong anion-exchange ProteinChips: potential use in diagnosis and prognosis. Proc Natl Acad Sci USA 2003, 100(21), 12343–12348.
- MacDonald TJ, Ladisch S. Antisense to integrin alpha v inhibits growth and induces apoptosis in medulloblastoma cells. *Antican*cer Res 2001, 21(6A), 3785–3791.
- Pomeroy SL, Tamayo P, Gaasenbeek M, Sturla LM, Angelo M, McLaughlin ME, et al. Prediction of central nervous system embryonal tumour outcome based on gene expression. Nature 2002, 415(6870), 436–442.
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, et al. Molecular portraits of human breast tumours. *Nature* 2000, 406(6797), 747–752.
- Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, et al. Repeated observation of breast tumour subtypes in independent gene expression data sets. Proc Natl Acad Sci USA 2003, 100(14), 8418–8423.
- van 't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M, et al. Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 2002, 415(6871), 530–536.
- 53. van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW, et al. A gene-expression signature as a predictor of survival in breast cancer. N Engl J Med 2002, 347(25), 1999–2009.
- West M, Blanchette C, Dressman H, Huang E, Ishida S, Spang R, et al. Predicting the clinical status of human breast cancer by using gene expression profiles. Proc Natl Acad Sci USA 2001, 98(20), 11462–11467.
- Hedenfalk I, Duggan D, Chen Y, Radmacher M, Bittner M, Simon R, et al. Gene-expression profiles in hereditary breast cancer. N Engl J Med 2001, 344(8), 539–548.
- 56. Gruvberger S, Ringner M, Chen Y, Panavally S, Saal LH, Borg A, et al. Estrogen receptor status in breast cancer is associated with remarkably distinct gene expression patterns. Cancer Res 2001, 61(16), 5979–5984.
- Alon U, Barkai N, Notterman DA, Gish K, Ybarra S, Mack D, et al. Broad patterns of gene expression revealed by clustering analysis of tumour and normal colon tissues probed by oligonucleotide arrays. Proc Natl Acad Sci USA 1999, 96(12), 6745–6750
- 58. Zou TT, Selaru FM, Xu Y, Shustova V, Yin J, Mori Y, et al. Application of cDNA microarrays to generate a molecular taxonomy capable of distinguishing between colon cancer and normal colon. Oncogene 2002, 21(31), 4855–4862.
- Hippo Y, Taniguchi H, Tsutsumi S, Machida N, Chong JM, Fukayama M, et al. Global gene expression analysis of gastric cancer by oligonucleotide microarrays. Cancer Res 2002, 62(1), 233–240.

- Takahashi M, Rhodes DR, Furge KA, Kanayama H, Kagawa S, Haab BB, et al. Gene expression profiling of clear cell renal cell carcinoma: gene identification and prognostic classification. Proc Natl Acad Sci USA 2001, 98(17), 9754–9759.
- Beer DG, Kardia SL, Huang CC, Giordano TJ, Levin AM, Misek DE, et al. Gene-expression profiles predict survival of patients with lung adenocarcinoma. Nat Med 2002, 8(8), 816–824.
- Garber ME, Troyanskaya OG, Schluens K, Petersen S, Thaesler Z, Pacyna-Gengelbach M, et al. Diversity of gene expression in adenocarcinoma of the lung. Proc Natl Acad Sci USA 2001, 98(24), 13784–13789.
- Bhattacharjee A, Richards WG, Staunton J, Li C, Monti S, Vasa P, et al. Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. Proc Natl Acad Sci USA 2001, 98(24), 13790–13795.
- 64. Singh R, Eeles RA, Durocher F, Simard J, Edwards S, Badzioch M, et al. High risk genes predisposing to prostate cancer development-do they exist? Prostate Cancer Prostatic Dis 2000, 3(4), 241–247.
- 65. LaTulippe E, Satagopan J, Smith A, Scher H, Scardino P, Reuter V, et al. Comprehensive gene expression analysis of prostate cancer reveals distinct transcriptional programs associated with metastatic disease. Cancer Res 2002, 62(15), 4499–4506.
- 66. Welsh JB, Sapinoso LM, Su AI, Kern SG, Wang-Rodriguez J, Moskaluk CA, et al. Analysis of gene expression identifies candidate markers and pharmacological targets in prostate cancer. Cancer Res 2001, 61(16), 5974–5978.
- Dhanasekaran SM, Barrette TR, Ghosh D, Shah R, Varambally S, Kurachi K, et al. Delineation of prognostic biomarkers in prostate cancer. Nature 2001, 412(6849), 822–826.
- Bittner M, Meltzer P, Chen Y, Jiang Y, Seftor E, Hendrix M, et al. Molecular classification of cutaneous malignant melanoma by gene expression profiling. Nature 2000, 406(6795), 536–540.
- Khan J, Wei JS, Ringner M, Saal LH, Ladanyi M, Westermann F, et al. Classification and diagnostic prediction of cancers using gene expression profiling and artificial neural networks. Nat Med 2001, 7(6), 673–679.
- Yeoh EJ, Ross ME, Shurtleff SA, Williams WK, Patel D, Mahfouz R, et al. Classification, subtype discovery, and prediction of outcome in pediatric acute lymphoblastic leukemia by gene expression profiling. Cancer Cell 2002, 1(2), 133–143.
- Hofmann WK, de Vos S, Elashoff D, Gschaidmeier H, Hoelzer D, Koeffler HP, et al. Relation between resistance of Philadelphiachromosome-positive acute lymphoblastic leukaemia to the tyrosine kinase inhibitor STI571 and gene-expression profiles: a gene-expression study. *Lancet* 2002, 359(9305), 481–486.
- Ferrando AA, Neuberg DS, Staunton J, Loh ML, Huard C, Raimondi SC, et al. Gene expression signatures define novel oncogenic pathways in T cell acute lymphoblastic leukemia. Cancer Cell 2002, 1(1), 75–87.
- Alizadeh AA, Staudt LM. Genomic-scale gene expression profiling of normal and malignant immune cells. *Curr Opin Immunol* 2000, 12(2), 219–225.
- 74. Shipp MA, Ross KN, Tamayo P, Weng AP, Kutok JL, Aguiar RC, et al. Diffuse large B-cell lymphoma outcome prediction by gene-expression profiling and supervised machine learning. Nat Med 2002, 8(1), 68–74.
- Rosenwald A, Wright G, Chan WC, Connors JM, Campo E, Fisher RI, et al. The use of molecular profiling to predict survival after chemotherapy for diffuse large-B-cell lymphoma. N Engl J Med 2002, 346(25), 1937–1947.
- Wong KK, Cheng RS, Mok SC. Identification of differentially expressed genes from ovarian cancer cells by MICROMAX cDNA microarray system. *Biotechniques* 2001, 30(3), 670–675.
- 77. Nishizuka S, Chen ST, Gwadry FG, Alexander J, Major SM, Scherf U, et al. Diagnostic markers that distinguish colon

- and ovarian adenocarcinomas: identification by genomic, proteomic, and tissue array profiling. *Cancer Res* 2003, **63**(17), 5243–5250.
- Shridhar V, Lee J, Pandita A, Iturria S, Avula R, Staub J, et al. Genetic analysis of early- versus late-stage ovarian tumours. Cancer Res 2001, 61(15), 5895–5904.
- Hough CD, Sherman-Baust CA, Pizer ES, Montz FJ, Im DD, Rosenshein NB, et al. Large-scale serial analysis of gene expression reveals genes differentially expressed in ovarian cancer. Cancer Res 2000, 60(22), 6281–6287.
- Wang K, Gan L, Jeffery E, Gayle M, Gown AM, Skelly M, et al. Monitoring gene expression profile changes in ovarian carcinomas using cDNA microarray. Gene 1999, 229(1–2), 101–108.
- Hough CD, Cho KR, Zonderman AB, Schwartz DR, Morin PJ. Coordinately up-regulated genes in ovarian cancer. *Cancer Res* 2001, 61(10), 3869–3876.
- Sawiris GP, Sherman-Baust CA, Becker KG, Cheadle C, Teichberg D, Morin PJ. Development of a highly specialized cDNA array for the study and diagnosis of epithelial ovarian cancer. *Cancer Res* 2002, 62(10), 2923–2928.
- Welsh JB, Sapinoso LM, Kern SG, Brown DA, Liu T, Bauskin AR, et al. Large-scale delineation of secreted protein biomarkers overexpressed in cancer tissue and serum. Proc Natl Acad Sci USA 2003, 100(6), 3410–3415.
- 84. Rangel LB, Sherman-Baust CA, Wernyj RP, Schwartz DR, Cho KR, Morin PJ. Characterization of novel human ovarian cancerspecific transcripts (HOSTs) identified by serial analysis of gene expression. *Oncogene* 2003, 22(46), 7225–7232.
- Ono K, Tanaka T, Tsunoda T, Kitahara O, Kihara C, Okamoto A, et al. Identification by cDNA microarray of genes involved in ovarian carcinogenesis. Cancer Res 2000, 60(18), 5007–5011.
- Kirchhoff C, Habben I, Ivell R, Krull N. A major human epididymis-specific cDNA encodes a protein with sequence homology to extracellular proteinase inhibitors. *Biol Reprod* 1991, 45(2), 350–357.
- 87. Schummer M, Ng WV, Bumgarner RE, Nelson PS, Schummer B, Bednarski DW, *et al.* Comparative hybridization of an array of 21,500 ovarian cDNAs for the discovery of genes overexpressed in ovarian carcinomas. *Gene* 1999, **238**(2), 375–385.
- Hellstrom I, Raycraft J, Hayden-Ledbetter M, Ledbetter JA, Schummer M, McIntosh M, et al. The HE4 (WFDC2) protein is a biomarker for ovarian carcinoma. Cancer Res 2003, 63(13), 3695– 3700.
- Yu JX, Chao L, Chao J. Prostasin is a novel human serine proteinase from seminal fluid. Purification, tissue distribution, and localization in prostate gland. *J Biol Chem* 1994, 269(29), 18843–18848.
- Diamandis EP, Yousef GM. Human tissue kallikreins: a family of new cancer biomarkers. Clin Chem 2002, 48(8), 1198–1205.
- 91. Yousef GM, Diamandis EP. Expanded human tissue kallikrein family—a novel panel of cancer biomarkers. *Tumour Biol* 2002, 23(3), 185–192.
- 92. Yousef GM, Diamandis EP. The new human tissue kallikrein gene family: structure, function, and association to disease. *Endocr Rev* 2001, **22**(2), 184–204.
- 93. Luo LY, Katsaros D, Scorilas A, Fracchioli S, Bellino R, van Gramberen M, et al. The serum concentration of human kallikrein 10 represents a novel biomarker for ovarian cancer diagnosis and prognosis. Cancer Res 2003, 63(4), 807–811.
- 94. Kishi T, Grass L, Soosaipillai A, Scorilas A, Harbeck N, Schmalfeldt B, et al. Human kallikrein 8, a novel biomarker for ovarian carcinoma. Cancer Res 2003, 63(11), 2771–2774.
- Diamandis EP, Yousef GM, Soosaipillai AR, Bunting P. Human kallikrein 6 (zyme/protease M/neurosin): a new serum biomarker of ovarian carcinoma. *Clin Biochem* 2000, 33(7), 579–583.
- 96. Frierson Jr HF, Moskaluk CA, Powell SM, Zhang H, Cerilli MH, Stoler MH, *et al.* Large-scale molecular and tissue microarray

- analysis of mesothelin expression in common human carcinomas. *Hum Pathol* 2003, **34**(6), 605–609.
- 97. Scholler N, Fu N, Yang Y, Ye Z, Goodman GE, Hellstrom KE, et al. Soluble member(s) of the mesothelin/megakaryocyte potentiating factor family are detectable in sera from patients with ovarian carcinoma. Proc Natl Acad Sci USA 1999, 96(20), 11531–11536.
- Huang G, Einstein M, Khabele D, Goldberg G. Serum biomarkers in epithelial ovarian cancer. Women's Oncol Rev 2003, 3(2), 117–122
- Xu Y, Shen Z, Wiper DW, Wu M, Morton RE, Elson P, et al. Lysophosphatidic acid as a potential biomarker for ovarian and other gynecologic cancers. JAMA 1998, 280(8), 719–723.
- 100. Morrison BH, Bauer JA, Kalvakolanu DV, Lindner DJ. Inositol hexakisphosphate kinase 2 mediates growth suppressive and apoptotic effects of interferon-beta in ovarian carcinoma cells. *J Biol Chem* 2001, 276(27), 24965–24970.

- Baker DL, Morrison P, Miller B, Riely CA, Tolley B, Westermann AM, et al. Plasma lysophosphatidic acid concentration and ovarian cancer. *JAMA* 2002, 287(23), 3081–3082.
- 102. Balkwill F, Bast RC, Berek J, Chenevix-Trench G, Gore M, Hamilton T, et al. Current research and treatment for epithelial ovarian cancer. A Position Paper from the Helene Harris Memorial Trust. Eur J Cancer 2003, 39(13), 1818–1827.
- 103. Gohagan JK, Prorok PC, Hayes RB, Kramer BS. The Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening Trial of the National Cancer Institute: history, organization, and status. Control Clin Trials 2000, 21(Suppl. 6), 251S–272S.
- 104. Menon U, Jacobs IJ. Ovarian cancer screening in the general population: current status. *Int J Gynecol Cancer* 2001, **1**(Suppl. 11), 3–6.
- 105. Carr K, Rosenblatt K, Petricoin E, Liotta L. Genomic and proteomic approaches to study human cancer: prospects for true patient-tailored therapy. *Hum Genomics* 2003, **1**(2), 32–38.
- 106. Conrads, 2003.